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<b>OXOID QUALITY ASSURANCE PRODUCT SPECIFICATION</b>		
<b>PSEUDOMONAS CN SELECTIVE SUPPLEMENT SR0102E</b>		

## PSEUDOMONAS CN SELECTIVE SUPPLEMENT

SR0102E

### Formula

Vial contents (each vial is sufficient to supplement 500ml of medium)

Cetrimide	100.0 mg
Sodium nalidixate	7.5 mg

### Description

A selective supplement for the isolation of *Pseudomonas* spp. when used with Pseudomonas Agar Base (CM0559).

### Directions

Aseptically add 2ml of 1:1 ethanol:sterile distilled water to 1 vial and mix gently to dissolve. Avoid frothing. Aseptically add the vial contents to 500ml of sterile Pseudomonas Basal Medium, prepared from Pseudomonas Agar Base (CM0559) plus 5ml glycerol and cooled to 50°C. Mix well and pour into sterile Petri dishes.

### Physical Characteristics

White pellet  
Sterility - passes test

### Microbiological Tests Using Optimum Inoculum Dilution

Control Medium: Tryptone Soya Agar

### Reactions after incubation at 37°C for 18 hours


Tested in Pseudomonas Agar Base CM0559

Medium is challenged with 10-100 colony-forming units

*Pseudomonas aeruginosa* ATCC® 25668

A satisfactory result is represented by recovery of positive strains equal to or greater than 70% of the control medium.

Some strains of *Pseudomonas aeruginosa* may produce little or no pigment with single colonies from low inocula.

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Medium is challenged with 1E+04 to 1E+06 colony-forming units

Inoculation using diminishing sweep technique

*Pseudomonas aeruginosa* ATCC® 25668 1-4mm straw colonies, green pigmentation

Medium is challenged with 1E+04 to 1E+06 colony-forming units

<i>Burkholderia cepacia</i>	ATCC® 25416	No growth or pinpoint to 0.25mm straw colonies
<i>Pseudomonas putida</i>	NCIMB 9034	No growth
<i>Pseudomonas fluorescens</i>	NCTC 3756	No growth
<i>Proteus hauseri</i>	ATCC® 13315	No growth
<i>Klebsiella pneumoniae</i>	ATCC® 29665	No growth
<i>Staphylococcus aureus</i>	ATCC® 25923	No growth
<i>Citrobacter freundii</i>	ATCC® 8090	No growth

Negative strains are inhibited or shall produce minimal growth

Additional challenging strains are employed.

#### Testing performed in accordance with ISO11133:2014

#### Reactions after incubation at 36 ± 2°C for 44 ± 4 hours

Tested with the addition of Pseudomonas CN Selective Supplement SR0102

Inoculation using membrane filtration technique

Medium is challenged with 50-120 colony forming units

<i>Pseudomonas aeruginosa</i>	ATCC® 27853	WDCM00025	1-4mm straw colonies, green pigmentation
<i>Pseudomonas aeruginosa</i>	ATCC® 9027	WDCM00026	1-4mm straw colonies, green pigmentation
<i>Pseudomonas aeruginosa</i>	ATCC® 10145	WDCM00024	1-4mm straw colonies, green pigmentation

A satisfactory result is represented by recovery of positive strains equal to or greater than 50% of the control medium. *Pseudomonas aeruginosa* should exhibit fluorescence under long wave UV light (365nm).

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Inoculation using diminishing sweep technique

Medium is challenged with 1E+04 to 1E+06 colony-forming units

<i>Escherichia coli</i>	ATCC® 8739	WDCM00012	No growth or 1-3mm cream colonies
<i>Escherichia coli</i>	ATCC® 25922	WDCM00013	No growth
<i>Enterococcus faecalis</i>	ATCC® 19433	WDCM00009	No growth
<i>Enterococcus faecalis</i>	ATCC® 29212	WDCM00087	No growth

A satisfactory result is represented by no growth or <50 cfu of *E.coli* ATCC®8739 WDCM00012 recovery of positive strains equal to or greater than 50% of the control medium. *Pseudomonas aeruginosa* should exhibit fluorescence under long wave UV light (365nm).

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### Revision History

Section / Step	Description of Change	Reason for Change	Reference
Creation of ISO11133 section	Update to include testing of ISO11133:2014	Change control	BT-CC-1341